ENZYMOLOGY

# Organometallic and radical intermediates reveal mechanism of diphthamide biosynthesis

Min Dong,[1] Venkatesan Kathiresan,[2] Michael K. Fenwick,[1] Andrew T. Torelli,[1] Yang Zhang,[1] Jonathan D. Caranto,[1] Boris Dzikovski,[1] Ajay Sharma,[2] Kyle M. Lancaster,[1] Jack H. Freed,[1] Steven E. Ealick,[1]* Brian M. Hoffman,[2]* Hening Lin[1,3]*

**Diphthamide biosynthesis involves a carbon-carbon bond-forming reaction catalyzed by a radical S-adenosylmethionine (SAM) enzyme that cleaves a carbon-sulfur (C–S) bond in SAM to generate a 3-amino-3-carboxypropyl (ACP) radical. Using rapid freezing, we have captured an organometallic intermediate with an iron-carbon (Fe–C) bond between ACP and the enzyme's [4Fe-4S] cluster. In the presence of the substrate protein, elongation factor 2, this intermediate converts to an organic radical, formed by addition of the ACP radical to a histidine side chain. Crystal structures of archaeal diphthamide biosynthetic radical SAM enzymes reveal that the carbon of the SAM C–S bond being cleaved is positioned near the unique cluster Fe, able to react with the cluster. Our results explain how selective C–S bond cleavage is achieved in this radical SAM enzyme.**

Diphthamide is a posttranslationally modified histidine residue on archaeal and eukaryotic translation elongation factor 2 (EF2), a protein essential for ribosomal protein synthesis (*1*–*3*). Diphthamide biosynthesis involves at least four steps and seven proteins (*4*). The first step of the biosynthesis is the transfer of a 3-amino-3-carboxypropyl (ACP) group from S-adenosyl-L-methionine (SAM) to a histidine residue of EF2 (Fig. 1). The enzyme that performs this reaction is a Dph2 (diphthamide biosynthesis protein 2) homodimer in archaea, such as *Pyrococcus horikoshii* (*Ph*Dph2) (*5*) or a Dph1-Dph2 heterodimer in eukaryotes (*6*). Dph2 binds an essential [4Fe-4S] cluster and is thought to use the reduced (1+) state of the cluster to cleave the $C_{\gamma,Met}$–S bond of SAM to generate a 3-amino-3-carboxypropyl (ACP) radical. The formation of the ACP radical is supported by the *Ph*Dph2-catalyzed generation of 2-aminobutyric acid and homocysteine sulfinic acid in the absence of the substrate protein, *Ph*EF2 (*5*). Additional support for a radical mechanism was provided by the reaction with a carboxyallyl SAM analog, $SAM_{CA}$ (*7*). However, no radical intermediate has been directly observed in the reaction with SAM itself, and the detailed reaction mechanism remains unknown.

Despite using the same components, a [4Fe-4S] cluster and SAM, *Ph*Dph2 is structurally unrelated to the much larger family of 5′-deoxyadenosine radical (5′-dA•) forming radical SAM (RS) enzymes. In those enzymes, the $C_{5',Ade}$–S bond of SAM is cleaved, generating a 5′-dA• that then initiates downstream reactions (Fig. 1) (*8*). The question of how the *Ph*Dph2 homodimer or eukaryotic Dph1-Dph2 bind SAM and achieve a different SAM cleavage pattern remains unanswered. Here, we report biochemical and spectroscopic studies of two kinetically competent intermediates that, together with x-ray crystal structures of archaeal Dph2 homodimers in complex with SAM, allow us to propose a reaction mechanism for this class of RS enzymes.

We used rapid freeze-quench (RFQ) to arrest reaction mixtures, which were then analyzed by electron paramagnetic resonance (EPR) and electron nuclear double-resonance (ENDOR) spectroscopies for radical intermediates. Because the optimum temperature of *Ph*Dph2 is above 60°C, we opted to use the yeast Dph1-Dph2 system, which is active at room temperature and thus more convenient for RFQ experiments. A solution containing Dph1-Dph2 and SAM was mixed with dithionite and freeze-quenched at 500 ms to 4 min. Each sample displayed a new EPR signal, with *g*-parallel value ($g_{||}$) = 2.036, *g*-perpendicular value ($g_{\perp}$)= 2.005 at 12 K (fig. S1). The intensity of this signal reached a maximum at 2 s quench time and decreased at longer delays (Fig. 2A). This species is not an ACP radical, which would not show such a high *g* shift and would exhibit resolvable $^{1}H$ hyperfine splittings from the methylene radical protons; nor is it an S-based radical, which could exhibit such *g* values but again would give resolved hyperfine $^{1}H$ splittings. Based on these observations and experience from previous work using $SAM_{CA}$ (*7*), we interpreted this new species as having an iron-sulfur cluster-based organometallic structure. Importantly, the EPR spectrum and *g* values of this species resemble those of the organometallic intermediate that was detected in the RS enzyme, pyruvate formate lyase-activating enzyme (PFL-AE) and assigned to an Fe-5′–C bond between the deoxyadenosyl group and the unique cluster iron (*9*). As confirmation, when this intermediate was generated with $^{57}Fe$-enriched enzyme, its EPR spectrum exhibited $^{57}Fe$-hyperfine line broadening (fig. S1). Thus, the reaction catalyzed by Dph1-Dph2 produces an organometallic intermediate. Its *g* values follow the pattern of a $[4Fe\text{-}4S]^{3+}$ cluster, $g_{||} > g_{\perp} > 2$ (*10*), where $g_{||}$ is sensitive to coordination at the unique Fe, suggesting a possible formal description of the intermediate as containing a carbanion bound to an oxidized cluster.

To establish the structure of this intermediate, we collected RFQ ENDOR samples prepared with (methionine-$^{13}C_5$)–SAM, in which the methionine carbons of SAM were uniformly labeled with $^{13}C$. Field-modulated continuous wave (CW) ENDOR spectra obtained at 2 K of this sample exhibits a $^{13}C$ doublet, with a $^{13}C$ hyperfine coupling constant of $A \approx 7.8$ MHz (Fig. 3A). This coupling constant is comparable to $A_{iso} = 9.4$ MHz observed for the Fe–[5′-$^{13}C$]–deoxyadenosyl bond

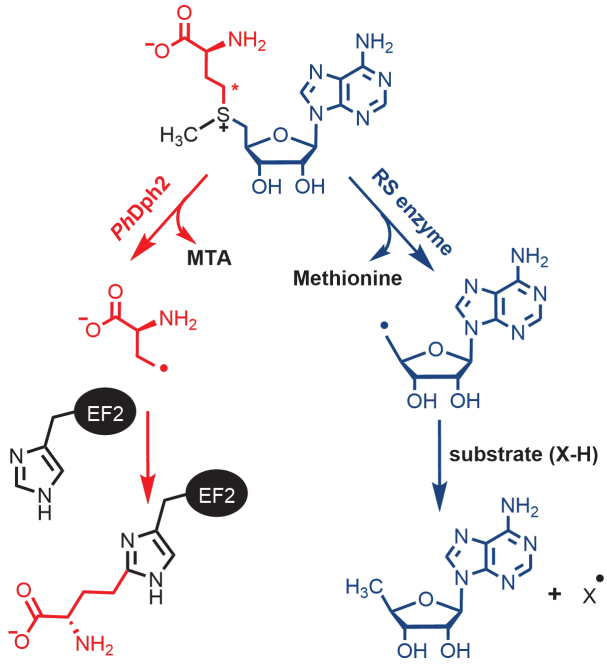


**Fig. 1. *Ph*Dph2 and 5′-dA•–forming radical SAM enzyme catalyzed reactions.** The red asterisk labels $C_{\gamma,Met}$ of SAM.

[1]Department of Chemistry and Chemical Biology, Cornell University, Ithaca, NY 14853, USA. [2]Department of Chemistry, Northwestern University, Evanston, IL 60208, USA. [3]Howard Hughes Medical Institute, Department of Chemistry and Chemical Biology, Cornell University, Ithaca, NY 14853, USA.
*Corresponding author. Email: see3@cornell.edu (S.E.E.); bmh@northwestern.edu (B.M.H.); hl379@cornell.edu (H.L.)

Downloaded from http://science.sciencemag.org/ on April 17, 2018

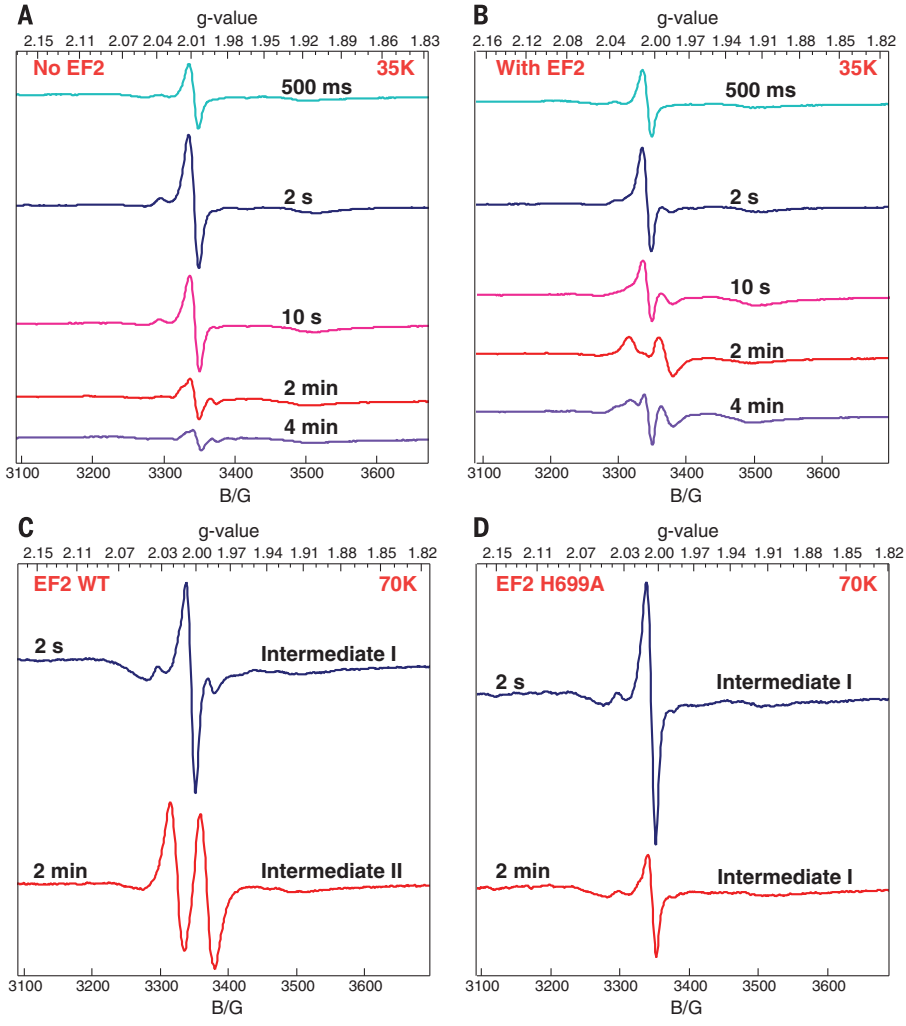


**Fig. 2. X-band CW EPR spectra of RFQ samples showing the formation of the organometallic intermediate I and organic radical intermediate II.** B/G (magnetic field/Gauss). (**A**) Reaction of reduced Dph1-Dph2 with SAM quenched at various time points ($T$ = 35 K). (**B**) Reaction of reduced Dph1-Dph2 with SAM and EF2 quenched at various time points ($T$ = 35 K). (**C**) Reaction of reduced Dph1-Dph2 with SAM and wild-type EF2 quenched at 2 s and 2 min ($T$ = 70 K). (**D**) Reaction of reduced Dph1-Dph2 with SAM and EF2 H699A mutant quenched at 2 s and 2 min ($T$ = 70 K). In early-time spectra, a variable contribution at $g$ = 2 from an unidentified organic radical enhances the intensity of the $g_{\perp}$ feature of intermediate I relative to that at $g_{||}$ and distorts its shape; fig. S1 presents spectra with little of the radical and thus a more faithful lineshape of intermediate I.

in the PFL-AE intermediate and together with the $^{57}$Fe hyperfine broadening provides strong support that this intermediate contains an Fe–C bond. However, in this case, the Fe-bonded carbon must originate from the $^{13}$C-labeled methionine, presumably the $C_{\gamma,Met}$.

In testing whether this organometallic species (denoted intermediate I) was an active reaction intermediate, we collected a RFQ EPR time course in the presence of the substrate protein, EF2. Intermediate I accumulated in the first 10 s (Fig. 2B). At later time points, the intermediate I signal diminished concomitantly with the appearance of a doublet radical signal centered at $g$ = 2 (Fig. 2B). This new signal, denoted intermediate II, reached a maximum at 2 min quench time and decreased at 4 min (Fig. 2B) in a process that ultimately leads to product formation.

The intermediate II EPR signal was distorted by saturation effect at 35 K (Fig. 2B) but was readily observed at 70 K (Fig. 2C), where spin relaxation is faster. Tentative assignment of the doublet splitting to coupling to a single proton with a large hyperfine splitting ($A \approx 120$ MHz) led us to hypothesize that this intermediate is an organic radical generated when the organometallic intermediate I reacted with the enzymatic target, histidine 699 (H699) of yeast EF2. To test this, we repeated the RFQ experiment with the EF2 His$^{699}$ to alanine mutant (H699A). With this EF2 mutant, only the organometallic intermediate I was detected at both 2 s and 2 min (Fig. 2D), and no intermediate II signal was detected. This experiment provides strong support that intermediate I is chemically competent.

To test the proposed structure, intermediate II was RFQ-trapped with (methionine-$^{13}C_5$)–SAM. The resulting X-band EPR spectrum showed additional splitting of $A_{iso}(^{13}C) \approx 59$ MHz and broadening compared with that of the natural-abundance SAM (Fig. 3B), demonstrating that the radical incorporates the ACP group of SAM. The EPR-resolved doublet, $A(^1H) \approx 120$ MHz, observed for intermediate II collapsed to a singlet when intermediate II was prepared with $^2H_5$-His–labeled EF2 protein (Fig. 3B), confirming that the splitting observed with the natural-abundance EF2 was associated with $^1$H of the histidine residue on EF2. This was further confirmed by $^1$H ENDOR of intermediate II with EF2, which exhibited a nonexchangeable $^1$H signal with the same coupling (fig. S2); an exchangeable coupling, $A(^1H) \approx 15$ MHz not resolved in the EPR spectrum was also seen. These results confirm that intermediate II is a radical produced by the reaction of an ACP radical from intermediate I with the imidazole ring of His$^{699}$ of the EF2 substrate.

Given the structure of the final diphthamide product, a candidate for the structure of intermediate II is an ACP-modified histidine radical, as shown in Fig. 4. The shape of the $^1$H doublet in the EPR spectrum (Fig. 3B) and the strongly coupled $^1$H ENDOR signal (fig. S2) suggest that the proton coupling is essentially isotropic, as expected for a proton β to the spin in the N-2p-π orbital (Fig. 4 and fig. S3) (*11*); the resolved $^{13}$C splitting in the EPR spectrum of intermediate II (Fig. 3B) prepared with (methionine-$^{13}C_5$)–SAM must come from the carbon of the ACP fragment bound to the His, again β to the spin site. The individual peaks of the $^1$H doublet are broad enough to contain unresolved $^{14}$N splitting introduced by the spin density on the $^{14}$Nδ of His (Fig. 4). As shown in fig. S4, the EPR spectrum of the natural-abundance radical is well simulated with a strong, nearly isotropic $^1$H coupling ($A_1 = A_2$ = 122 MHz; $A_3$ = 103 MHz) and a highly anisotropic $^{14}$N

Downloaded from http://science.sciencemag.org/ on April 17, 2018

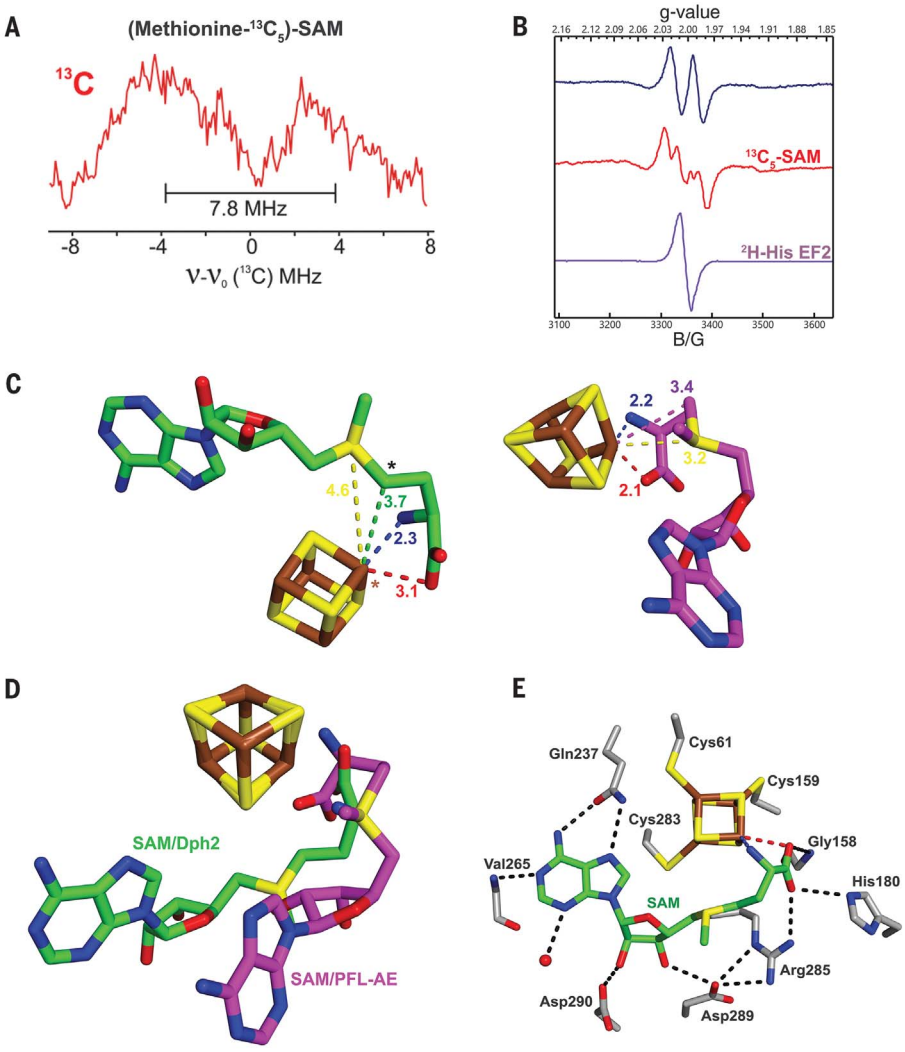


**Fig. 3. EPR and ENDOR spectra of isotope-labeled intermediates and crystal structures of SAM-bound *Cmn*Dph2.** (**A**) 35 GHz CW $^{13}$C ENDOR spectra for intermediate I with (methionine-$^{13}C_5$)–SAM. (**B**) Isotope EPR study of the organic radical intermediate II on EF2 quenched at 2 min ($T$ = 70 K). Intensity adjusted for comparison. (**C**) Cluster and bound SAM in the structure of *Cmn*Dph2 (left), in comparison with cluster and bound SAM in the structure of PFL-AE (right, PDB 3CB8) (*13*). Distances are given in angstroms. The black and brown asterisks label $C_{\gamma,Met}$ of SAM and the differentiated iron of the [4Fe-4S] cluster, respectively. (**D**) Overlay of SAM and cluster in *Cmn*Dph2 and PFL-AE structures. (**E**) *Cmn*Dph2 active site showing the binding of SAM. Hydrophobic interactions with purine ring of SAM are not shown for clarity.

hyperfine coupling ($A_1 = A_2 = 4$ MHz, $A_3 = 54$ MHz). $^{15}$N features seen in the ENDOR of the $^{15}$N-His EF2 sample (fig. S5) likely are a superposition of signals associated with the $A_1/A_2$ components of this histidine nitrogen (assigned as Nδ in Fig. 4), and those from the less strongly coupled His $^{15}$Nε. Finally, weakly coupled $^{13}$C ENDOR signals seen when intermediate II was prepared with $^{13}$C-His EF2 (figs. S5 and S6) are assigned to His carbons bonded to the spin-bearing Nδ (Fig. 4) and account for the slight broadening in the components of the $^1$H doublet in this isotopologue; an exchangeable $^1$H seen in ENDOR (fig. S2) is thought to reside on Nε of His.

The time scale for intermediate II formation measured by EPR tracked that of intermediate I decay, whereas the reaction product, ACP-modified EF2 as quantified by mass spectrometry, accumulates on the same time scale as the intermediate II decay (fig. S7). These results indicate that both intermediate I and intermediate II are formed in a kinetically competent fashion. Thus, we have trapped two sequential intermediates, an organometallic intermediate I, which gives rise to an organic radical intermediate II, which goes on to complete the first step of diphthamide biosynthesis.

To gain further understanding into the reaction mechanism, we obtained crystal structures of the RS enzymes involved in diphthamide biosynthesis in complex with SAM. A 2.3-Å resolution structure of [4Fe-4S]$^{2+}$–loaded *Ph*Dph2 in complex with SAM showed some evidence of SAM cleavage. A 2.1-Å resolution structure of *Candidatus Methanoperedens nitroreducens* Dph2 (*Cmn*Dph2) showed mostly uncleaved SAM. Both structures showed average N–Fe and O–Fe distances of 2.3 and 3.1 Å, respectively (Fig. 3C and fig. S8), suggesting that, similar to 5′-dA•–forming RS enzymes, the amino and carboxylate groups of SAM coordinate the unique Fe of the [4Fe-4S] cluster in Dph2. This is consistent with our recent study showing that both the amino and carboxylate of SAM are important for the *Ph*Dph2 and Dph1-Dph2–catalyzed reactions (*12*).

However, SAM in both *Ph*Dph2 and *Cmn*Dph2 binds with a distinct geometry compared with that in 5′-dA•–forming RS enzymes such as PFL-AE (*13*) (Fig. 3D). We further obtained additional crystal structures using both *Ph*Dph2 and *Cmn*Dph2 that support the geometry shown in Fig. 3, C and D (see figs. S9 to S11). The SAM binding site of *Cmn*Dph2 is shown in Fig. 3E. Gln$^{237}$ forms hydrogen bonds with N6 and N7 of the adenine ring and the amide nitrogen of Val$^{265}$ hydrogen bonds to N1. The adenine ring is sandwiched between Phe$^{58}$ and Ile$^{286}$. Asp$^{289}$ and Asp$^{290}$ form hydrogen bonds with the hydroxyl groups of the ribosyl moiety, whereas Arg$^{285}$, His$^{180}$, and Gly$^{158}$ form hydrogen bonds with the carboxylate of SAM.

Most strikingly, the $C_{\gamma,Met}$ distance to the unique Fe, as averaged over the *Ph*Dph2 and *Cmn*Dph2 structures, is 3.7 Å, which is closer than $S_{Met}$ to the Fe (the $S_{Met}$–Fe distance is 4.6 Å). In contrast, in the PFL-AE structure, the $S_{Met}$ is closer to the unique Fe than $C_{5',Ade}$, with an $S_{Met}$–Fe distance of 3.2 Å (Fig. 3C). The observation that in *Ph*Dph2 and *Cmn*Dph2, $C_{\gamma,Met}$ was closer than $S_{Met}$ to the unique Fe suggests that during the $C_{\gamma,Met}$–$S_{Met}$ bond cleavage, the electron is transferred from the Fe–S cluster via $C_{\gamma,Met}$, whereas for PFL-AE (and generally believed for all 5′-dA•–forming RS enzymes), the electron is transferred from the Fe–S cluster via $S_{Met}$ (*8*).

Based on the two intermediates that we have detected and the structures of *Ph*Dph2 and *Cmn*Dph2 in complex with SAM, we propose a reaction mechanism for the first step of

Downloaded from http://science.sciencemag.org/ on April 17, 2018

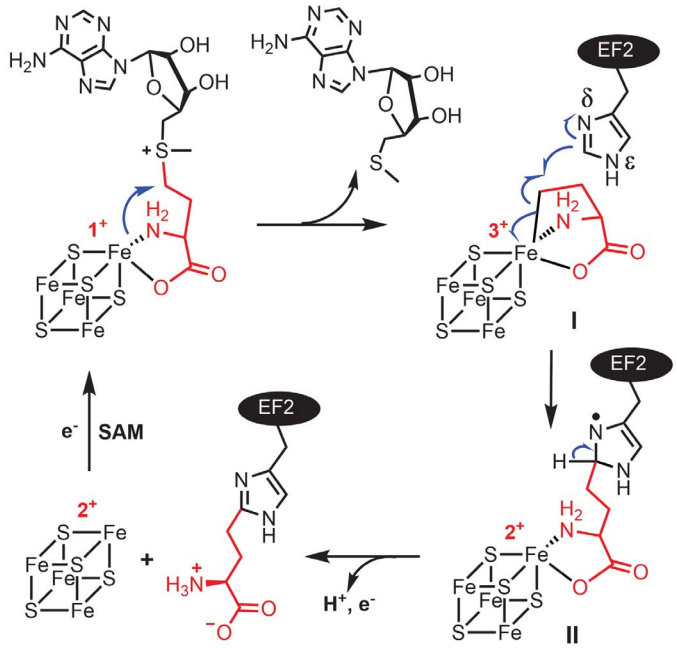


**Fig. 4. The proposed reaction mechanism of diphthamide biosynthetic RS enzymes.**

diphthamide biosynthesis (Fig. 4). The unique iron of a $[4Fe\text{-}4S]^{+}$ cluster in an archaeal Dph2 homodimer or eukaryotic Dph1-Dph2 heterodimer attacks the γ carbon of the methionine in SAM, generating a 3-amino-3-carboxypropyl-$[4Fe\text{-}4S]^{3+}$ organometallic intermediate I and 5′-methylthioadenosine (MTA). The organometallic intermediate I is essentially a stabilized ACP radical. In the presence of the substrate EF2, the Fe–C bond can break homolytically, allowing the ACP radical to react with the imidazole ring of the histidine residue of EF2 and generate the organic radical, intermediate II. Intermediate II then loses a proton and an electron to form an ACP-modified histidine.

In the study of PFL-AE, it was noted that the Fe-[5′-C]-deoxyadenosyl organometallic intermediate could be generated through a radical reaction or nucleophilic reaction (*9*). In our case, the formation of intermediate I could occur via homolytic reductive cleavage of SAM to form an ACP radical, which then reacts with the unique Fe (stepwise one-electron transfer) to form the organometallic intermediate I. Alternatively, a concerted one-step, two-electron transfer (nucleophilic attack on $C_{\gamma,Met}$) can form intermediate I directly. The fact that $C_{\gamma,Met}$ was only 3.7 Å away from the unique Fe, whereas the sulfonium moiety was 4.6 Å away (Fig. 3C), favors the latter mechanism, although we cannot eliminate the homolytic cleavage mechanism.

The structure also provides a simple solution to the question, how does the archaeal Dph2 homodimer or eukaryotic Dph1-Dph2 heterodimer cleave a different C–S bond in SAM compared with all 5′-dA•–forming RS enzymes? If the cluster interacts with and transfers electrons to $C_{\gamma,Met}$ instead of $S_{Met}$, the only C–S bond that can be broken in SAM is the $C_{\gamma,Met}$–S bond. Previously, Kampmeier proposed that a stereo-electronic control mechanism could explain the different cleavage patterns (*14*). This model rationalizes the C–S bond cleavage in RS enzymes using a radical displacement reaction, with the formation of an Fe–S bond accompanying the cleavage of the C–S bond (fig. S12). This radical displacement reaction also requires that the C–S–Fe atoms involved in the release of a radical on carbon are arranged roughly colinearly. Thus, if SAM in archaeal Dph2 homodimer or eukaryotic Dph1-Dph2 is bound in a conformation with $S_{Met}$ close to the Fe–S cluster and colinear $C_{\gamma,Met}$–S–Fe arrangement, it would lead to the cleavage of $C_{\gamma,Met}$–S. However, our structure indicates that in archaeal Dph2 homodimer, the $C_{\gamma,Met}$ is close to the unique Fe. The $C_{\gamma,Met}$ mediates electron transfer from the Fe–S cluster, dictating which bond is cleaved. This is an important difference between diphthamide biosynthetic RS enzymes and 5′-dA•–forming RS enzymes. Why does nature choose this way to control the bond cleavage in diphthamide biosynthetic RS enzymes? Perhaps conformational constraints on SAM prevent the $C_{\gamma,Met}$–S–Fe colinear arrangement proposed by Kampmeier, whereas the roughly colinear S–$C_{\gamma,Met}$–Fe arrangement as revealed by the structures is feasible (fig. S12).

In summary, this study has explained how diphthamide biosynthetic RS enzymes create the EF2 product through cleavage of the $C_{\gamma,Met}$–S bond of SAM in a process that involves first an organometallic intermediate, then a radical intermediate that collapses to product. Together with the recent study showing an organometallic intermediate in a 5′-dA•–forming RS enzyme PFL-AE, these studies further suggest that an organometallic intermediate may serve as a stabilized form of the highly reactive primary organic radical and provide a strong parallel between RS enzymes and adenosylcobalamin-dependent enzymes.

## REFERENCES AND NOTES


1. E. A. Robinson, O. Henriksen, E. S. Maxwell, *J. Biol. Chem.* **249**, 5088–5093 (1974).
2. B. G. Van Ness, J. B. Howard, J. W. Bodley, *J. Biol. Chem.* **255**, 10717–10720 (1980).
3. B. G. Van Ness, J. B. Howard, J. W. Bodley, *J. Biol. Chem.* **255**, 10710–10716 (1980).
4. R. Schaffrath, W. Abdel-Fattah, R. Klassen, M. J. Stark, *Mol. Microbiol.* **94**, 1213–1226 (2014).
5. Y. Zhang *et al.*, *Nature* **465**, 891–896 (2010).
6. M. Dong *et al.*, *J. Am. Chem. Soc.* **136**, 1754–1757 (2014).
7. M. Dong *et al.*, *J. Am. Chem. Soc.* **138**, 9755–9758 (2016).
8. J. B. Broderick, B. R. Duffus, K. S. Duschene, E. M. Shepard, *Chem. Rev.* **114**, 4229–4317 (2014).
9. M. Horitani *et al.*, *Science* **352**, 822–825 (2016).
10. G. E. Cutsail 3rd, J. Telser, B. M. Hoffman, *Biochim. Biophys. Acta* **1853**, 1370–1394 (2015).
11. A. Carrington, A. D. McLachlan, *Introduction to Magnetic Resonance with Applications to Chemistry and Chemical Physics* (Harper & Row, New York, 1967).
12. M. Dong *et al.*, *J. Am. Chem. Soc.* **139**, 5680–5683 (2017).
13. J. L. Vey *et al.*, *Proc. Natl. Acad. Sci. U.S.A.* **105**, 16137–16141 (2008).
14. J. A. Kampmeier, *Biochemistry* **49**, 10770–10772 (2010).


## ACKNOWLEDGMENTS


We thank M. Luo at Memorial Sloan–Kettering Cancer Center for providing the methionine adenosyltransferase I plasmid. We thank P. Doan for help in confirming the organometallic nature of intermediate I and M. Oliver Ross for assistance in collecting ENDOR spectra. We thank S. Zhang (Proteomics and Metabolomics Facility at Cornell) for help with the mass spectrometry experiment. M. D. thanks T. L. Grove and R. D. Britt for helpful discussion. **Funding**: This work was supported by grants from the National Institutes of Health National Institute of General Medical Sciences (GM088276 to H.L., P41GM103521 to J.H.F., P41GM103403 to S.E.E, GM 111097 to B.M.H., and GM124908 to K.M.L.). This work is based on research conducted at the Northeastern Collaborative Access Team beamlines. The Pilatus 6M detector on 24-ID-C beamline is funded by a NIH-ORIP HEI grant (S10 RR029205). This research used resources of the Advanced Photon Source, a U.S. Department of Energy (DOE) Office of Science User Facility operated for the DOE Office of Science by Argonne National Laboratory under contract no. DE-AC02-06CH11357. **Author contributions**: M.D. contributed to experimental design, performed the biochemical experiments, and prepared the proteins for spectroscopic studies. V.K. and A.S. performed the ENDOR studies. M.K.F., A.T.T., and Y.Z. carried out the crystallization and structure determination of all the proteins. J.D.C. and K.M.L. provided critical support for the RFQ experiments and EPR simulation and performed the fitting of kinetic data. M.D., B.D., and J.H.F performed the X-band EPR measurements. S.E.E. directed the x-ray structural studies; B.M.H. directed the ENDOR studies and contributed to experimental design; H.L. directed the biochemical studies, contributed to experimental design, and coordinated collaborations. M.D., H.L., B.M.H., and S.E.E. wrote the manuscript with input from V.K., M.K.F., A.T.T., Y.Z., and J.D.C. All authors reviewed, edited, and approved the manuscript. **Competing interests**: The authors declare no competing financial interests. **Data and materials availability**: Crystallographic models have been deposited in the Protein Data Bank under accession codes 6BXK for *Ph*Dph2/MTA, 6BXL for *Ph*Dph2/SAM, 6BXM for *Cmn*Dph2/SAM1, 6BXN for *Cmn*Dph2/SAM2, and 6BXO for *Cmn*Dph2/SAH. Raw data and materials associated with the main text and the supplementary materials are available for sharing upon request.


## SUPPLEMENTARY MATERIALS

www.sciencemag.org/content/359/6381/1247/suppl/DC1
Materials and Methods
Supplementary Text
Figs. S1 to S12
Table S1
References (*15*–*35*)

12 August 2017; accepted 30 January 2018
10.1126/science.aao6595

Downloaded from http://science.sciencemag.org/ on April 17, 2018

Science

# Organometallic and radical intermediates reveal mechanism of diphthamide biosynthesis

Min Dong, Venkatesan Kathiresan, Michael K. Fenwick, Andrew T. Torelli, Yang Zhang, Jonathan D. Caranto, Boris Dzikovski, Ajay Sharma, Kyle M. Lancaster, Jack H. Freed, Steven E. Ealick, Brian M. Hoffman and Hening Lin

*Science* **359** (6381), 1247-1250.
DOI: 10.1126/science.aao6595

**A quick freeze shows an enzyme's secrets**

Organic radicals are chemically useful in enzymatic reactions but are often hard to observe, owing to their short lifetimes. Dong *et al.* used rapid freeze-quench methods to trap two intermediates formed by a noncanonical radical S-adenosylmethionine (SAM) enzyme: a fragmented SAM molecule bound to the iron-sulfur cluster through an iron-carbon bond and a product-like radical. The structure of the SAM-bound enzyme reveals a noncolinear arrangement of carbon, sulfur, and iron atoms. The arrangement of bonds suggests that the organometallic intermediate may be created through a two-electron nucleophilic mechanism. A subsequent radical intermediate is formed on the protein substrate and resolves by oxidation to form the amino acid product diphthamide.

*Science*, this issue p. 1247

**ARTICLE TOOLS** http://science.sciencemag.org/content/359/6381/1247

**SUPPLEMENTARY MATERIALS** http://science.sciencemag.org/content/suppl/2018/03/14/359.6381.1247.DC1

**REFERENCES** This article cites 34 articles, 8 of which you can access for free
http://science.sciencemag.org/content/359/6381/1247#BIBL

**PERMISSIONS** http://www.sciencemag.org/help/reprints-and-permissions

Use of this article is subject to the Terms of Service

*Science* (print ISSN 0036-8075; online ISSN 1095-9203) is published by the American Association for the Advancement of Science, 1200 New York Avenue NW, Washington, DC 20005. 2017 © The Authors, some rights reserved; exclusive licensee American Association for the Advancement of Science. No claim to original U.S. Government Works. The title *Science* is a registered trademark of AAAS.

Downloaded from http://science.sciencemag.org/ on April 17, 2018